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Allicin from garlic inhibits the biofilm formation and urease activity of *Proteus mirabilis* in vitro

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One sentence summary: Allicin from garlic inhibits the growth, biofilm formation and urease activity of *Proteus mirabilis* in vitro.

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ABSTRACT

Several virulence factors contribute to the pathogenesis of *Proteus mirabilis*. This study determined the inhibitory effects of allicin on urease, hemolysin and biofilm of *P. mirabilis* ATCC 12453 and its antimicrobial activity against 20 clinical isolates of *P. mirabilis*. Allicin did not inhibit hemolysin, whereas it did inhibit relative urease activity in both pre-lysed (half-maximum inhibitory concentration, IC₅₀ = 4.15 µg) and intact cells (IC₅₀ = 21 µg) in a concentration-dependent manner. Allicin at sub-minimum inhibitory concentrations (2–32 µg mL⁻¹) showed no significant effects on the growth of the bacteria (*P* > 0.05), but it reduced biofilm development in a concentration-dependent manner (*P* < 0.001). A higher concentration of allicin was needed to inhibit the established biofilms. Using the microdilution technique, the MIC₉₀ and MBC₉₀ values of allicin against *P. mirabilis* isolates were determined to be 128 and 512 µg mL⁻¹, respectively. The results suggest that allicin could have clinical applications in controlling *P. mirabilis* infections.

Keywords: allicin; *Proteus mirabilis*; antimicrobial; urease; hemolysin; biofilm

INTRODUCTION

Proteus mirabilis, a member of the Enterobacteriaceae family, is one of the leading causes of urinary tract infections (UTIs) in patients with indwelling catheters or structural abnormalities in the urinary tract (Coker *et al.* 2000). *Proteus mirabilis* UTIs are most commonly associated with urinary tract obstruction, blockage of urinary catheters, bladder and kidney stone formation, and

bacteriuria. Intrinsically, *P. mirabilis* UTIs are often persistent and difficult to treat (Rozalski, Sidorczyk and Kotelko 1997). Furthermore, the development of extended spectrum beta-lactamase producing and multiple drug resistant strains of *P. mirabilis* (Mokracka, Gruszczynska and Kaznowski 2012; Kurihara *et al.* 2013) have made treating *P. mirabilis* infections more difficult. Therefore, finding antibiotics with new modes of action and alternatives to commonly used antimicrobial therapies for

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